

# PETER BRAZDA

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## EDUCATION

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University of Debrecen, Ph.D., Cell Biology and Biophysics 9/2005 – 3/2015  
University of Debrecen, M.Sc., Biotechnology 9/1999 – 7/2005

## RESEARCH EXPERIENCE

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### **Princess Máxima Center for Pediatric Oncology (PMC), Utrecht, The Netherlands**

*Postdoctoral research* with Professor Henk Stunnenberg 6/2019 – present

- Project 1: Transcriptome profiling in Pheochromocytoma at single cell level
- Project 2: Transcriptome profiling in pediatric AML at single cell level

### **Radboud University, Institute for Molecular Life Sciences (RIMLS), Nijmegen, The Netherlands**

*Postdoctoral research* with Professor Henk Stunnenberg 6/2017 – 6/2019

- Project 1: Transcriptome profiling in Pheochromocytoma at single cell level
- Project 2: Transcriptomic analysis of early innate immune events after pertussis booster vaccination

### **Delft University of Technology, Department of Bionanoscience, Delft, The Netherlands**

*Postdoctoral research* with Professor Nynke Dekker 4/2015 – 5/2017

- Project 1: Accessory helicase (UvrD) action in *E. coli* replication-transcription conflicts with single molecule microscopy
- Project 2: The dynamics of *E. coli* replication termination (Tus) with super-resolution microscopy and large population genomics

### **German Cancer Research Center (DKFZ), Biophysics of Macromolecules Division, Heidelberg, Germany**

*Guest researcher* with Professor Jorg Langowski 4/2013 – 12/2014

- Several visits to test and carry out FCS, FCCS (fluorescence (cross-)correlation spectroscopy) and also single-plane illumination microscopy (SPIM-FCS) measurements on the setup designed and built by the group.

### **University of Debrecen, Department of Biochemistry and Molecular Biology, Debrecen, Hungary**

*Graduate research* with Professor Laszlo Nagy and Dr Gyorgy Vamosi 9/2005 – 3/2015

- Thesis: Determination of dynamic properties of nuclear receptors
- Project 1: The mobility of retinoic acid receptor (RAR) during activation by fluorescence correlation spectroscopy (FCS)
- Project 2: The dynamic properties of retinoic-x receptor (RXR) during activation, chromatin association and coregulator binding as seen by single-plane illumination FCS (SPIM-FCS) and large population genomics (ChIP-seq)

- Project 3: The ligand-dependent dimerization (homo- and heterodimer) of RXR investigated by flow cytometry and single cell imaging

*Undergraduate research* with Professor Laszlo Nagy

12/2002 – 9/2005

- Thesis: Investigation of a novel retinoid-x receptor antagonist ligand (Best presentation in Natural Sciences at the National Conference of Scientific Students' Association (OTDK))
- Project 1: The study of a novel ligand on the dimerization- and transactivation ability of RXR with transient transfection assay

**PUBLICATIONS:** <http://tiny.cc/dldh5y>

## **LEADERSHIP EXPERIENCE**

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- International Genetically Engineered Machines (iGEM), tutor for the Debrecen team 2010-Boston, 2011-Amsterdam
- Researchers' Night, organizer at the University of Debrecen, 2008, 2009, 2011, 2012
- South-East European (SEE) Science Festival, organizer of local events, 2013
- Association of Biologist Students of the University of Debrecen, president, 2007

## **TEACHING ACTIVITIES**

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- Practical in Biochemistry and Molecular Biology for medical students (Uni. of Debr.)
- Seminar in Molecular Biology for medical students. (Uni. of Debr.)
- Co-tutury support of 2 undergraduate students and support for 7 more students (Uni. of Debr.)
- Co-tutury support of 2 undergraduate students (TU Delft, RIMLS)

## **RESEARCH-RELATED SKILLS**

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**RNA:** isolation, RNA sequencing (bulk, single cell: CELseq2, 10x), scATACseq, RT-qPCR

**DNA:** isolation, molecular cloning, mutagenesis, chromatin IP

**NGS:** performing sequencing runs on Illumina (MiniSeq, Novaseq) platforms

**cell culturing:** cell-line related tasks, transfection, mammalian two-hybrid assay, bacterial strain engineering (recombination, transduction), processing of primary tissue samples

**analysis:** flow cytometry, confocal microscopy (imaging, FCS (fluorescence (cross)correlation spectroscopy, FRAP, FRET), fluorescence microscopy (wide field, super-resolution), single cell RNAseq data analysis, R-language